

Chapter 9: Insects for Biocontrol of Aquatic Weeds

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Introduction

Biocontrol of aquatic weeds with insects has resulted in the successful establishment of many potential biocontrol insects since it was first attempted in the US against alligatorweed in 1964. Aquatic weeds have historically been a more serious problem in the southern US due to the moderate climate and shallow lakes in these regions where weeds often cover large areas. Consequently, the greatest body of research on biocontrol has focused on weeds of the southern US. The following section describes in detail the relationship of particular biocontrol insects introduced into the US and their target weeds.

Alligatorweed (*Alternanthera philoxeroides*)

(see <http://el.erdc.usace.army.mil/aqua/APIS/apishelp.htm> for more information)

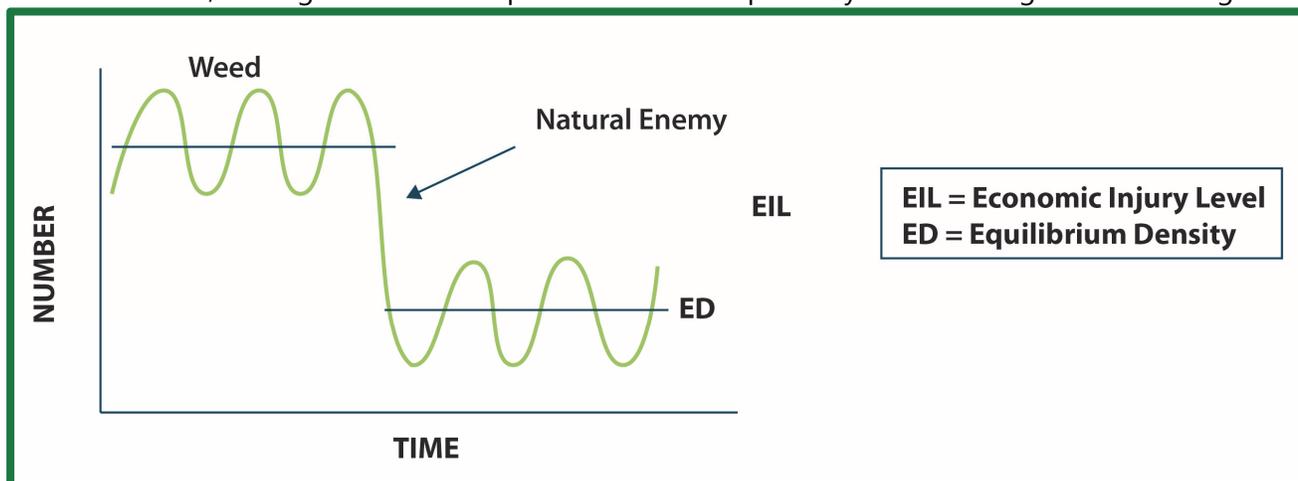
Enemy	Type	Origin (date)	Success	Comments
<i>Agasicles hygrophila</i>	Beetle	Argentina (1964)	Complete (south); Negligible (north)	Found throughout the southern 2/3 of the range of alligatorweed in the US where it provides almost complete control
<i>Amynothrips andersoni</i>	Thrips	Argentina (1967)	Negligible	Attacks terrestrial plants more than the other species
<i>Arcola (=Vogtia) malloi</i>	Moth	Argentina (1971)	Negligible	Most important control agent in the upper Mississippi valley

If this manual had been written in the 1960s and 1970s, alligatorweed (introduced in the late 1800s) would have been included as one of the worst weeds in the US in Chapter 15. The alligatorweed flea beetle (*Agasicles hygrophila*) was introduced in 1964 and has provided excellent control of the floating form of alligatorweed from southern Florida along the Gulf Coast to southern Texas. Unfortunately the alligatorweed flea beetle is not as cold-tolerant as alligatorweed and insect populations die out during severe winters in the central and northern parts of the Gulf states.

Alligatorweed remains a problem in areas such as central and northern Texas, Mississippi, Alabama, Georgia and the Carolinas. The alligatorweed flea beetle is self-sustaining in its southern range but not in the north. The US Army Corps of Engineers periodically collects and re-releases



the beetle in northern areas during spring to reestablish northern populations. This is an example of combining augmentation with classical biocontrol. The alligatorweed flea beetle has eliminated the need for other forms of control in natural areas when it is well-established. The *Amynothrips* and *Arcola* insects also are established on alligatorweed; the *Amynothrips* attacks the terrestrial alligatorweed plants more than do the other species. However, control of alligatorweed is largely attributed to the alligatorweed flea beetle. Alligatorweed provides a good example of how a biocontrol agent controls its weedy host plant without completely eradicating the population of the weed. Alligatorweed grows quickly in spring and populations of the alligatorweed flea beetle increase as well, but lag behind development of the host plant. By the time alligatorweed has grown



enough to become problematic, the population of the alligatorweed flea beetle reaches a density sufficient to destroy most of the alligatorweed. The number of alligatorweed flea beetles then decreases, alligatorweed growth resumes and the cycle begins anew. This is a nearly perfect example of a highly successful insect biocontrol program that adequately controls an invasive aquatic plant. Furthermore, 40 to 50 years after their introduction, none of the three insects released to control alligatorweed have been found feeding on, reproducing on or otherwise affecting nontarget native species.

Waterhyacinth (Chapter 15.7)

(see <http://el.erc.usace.army.mil/aqua/APIS/apishelp.htm> for more information)

Enemy	Type	Origin (date)	Success	Comments
<i>Neochetina bruchi</i>	Weevil	Argentina (1974)	Substantial	Widely distributed throughout the range of waterhyacinth in the US
<i>Neochetina eichhorniae</i>	Weevil	Argentina (1972)	Substantial	
<i>Niphograptus albiguttalis</i>	Moth	Argentina (1977)	Negligible	Prefers plants with short bulbous petioles
<i>Orthogalumna terebrantis</i>	Mite	USA (native)	Negligible	Produces characteristic dark stripes in the leaves; also attacks pickerelweed
<i>Megamelus scutellaris</i>	Bug	South America (2010)	Established	

Two *Neochetina* weevils and the *Niphograptus* stem-boring caterpillar have been released as biocontrol agents of waterhyacinth. The life cycle of the *Neochetina* weevils requires about 2 to 3 months to complete and is dependent on temperature. These weevils act on waterhyacinth by causing feeding damage that reduces the plant's ability to regenerate. Adult weevils produce

characteristic rectangular feeding scars on the leaves, whereas larvae tunnel inside the leaf petioles to the crown or meristem where they damage new growth. Feeding damage also allows plant pathogens to invade the feeding scars and larval tunnels, which further weakens the plant. The life cycle of the *Niphograpta* caterpillar is completed in about 4 to 5 weeks. This insect prefers to attack smaller plants with bulbous petioles; petioles that are attacked often become waterlogged and die. However, the impact of the *Niphograpta* caterpillar has been difficult to evaluate because it causes tremendous damage for only a brief period and then disappears. The *Neochetina* weevils and the *Niphograpta* stem-boring caterpillar are established and occur almost everywhere waterhyacinth is distributed throughout the southern US. Growth of waterhyacinth is suppressed and vegetative reproduction is reduced, but other means of control are necessary in most areas.

Hydrilla (Chapter 15.1)

(see <http://el.ercd.usace.army.mil/aqua/APIS/apishelp.htm> for more information)

Enemy	Type	Origin (date)	Success	Comments
<i>Bagous affinis</i>	Weevil	India (1987)	Not Established	
<i>Bagous hydrillae</i>	Weevil	Australia (1991)	Not Established	
<i>Cricotopus lebetis</i>	Midge	Unknown (adventive)	Negligible?	Damages growing tips of hydrilla Also see http://edis.ifas.ufl.edu/IN211
<i>Hydrellia balciunasi</i>	Fly	Australia (1989)	Negligible	Found primarily in Texas
<i>Hydrellia pakistanae</i>	Fly	India (1987)	Negligible?	Widely distributed on dioecious hydrilla in the southeastern and south-central US
<i>Parapoynx diminutalis</i>	Moth	Asia (adventive)	Negligible	Causes localized occasional heavy damage to hydrilla
<i>Ctenopharyngodon idella</i>	Fish	China (1963)	Substantial	Throughout the US by permit (Chapter 9) Also see http://plants.ifas.ufl.edu/guide/grasscarp.html

Two *Bagous* weevils (one from India that attacks tubers and one from Australia that mines stems) have been introduced as biocontrol agents for hydrilla, but both have failed to establish. However, two *Hydrellia* flies (one from India and one from Australia) have become established. The fly *H. pakistanae* is widespread in the southern US, whereas *H. balciunasi* is localized in distribution. Populations of *Hydrellia* flies have not reached densities high enough to control hydrilla, possibly due to parasitism of the pupae by a native wasp or perhaps other environmental factors. The entire life cycle for both flies is completed in about 3 to 4 weeks, which should allow development of high insect populations. The adventive *Parapoynx* moth from Asia probably entered the US via the aquarium trade and was discovered in Florida feeding on hydrilla in 1976. The life cycle of *Parapoynx* is completed in 4 to 5 weeks; the moth was never studied or approved for release, but large populations of hydrilla are occasionally completely defoliated by the moth. The adventive naturalized nonnative *Cricotopus* midge has been associated with hydrilla declines in several Florida locations since 1992. The life cycle of *Cricotopus* is completed in 1 to 2 weeks and developing larvae of the midge mine the shoot tips of hydrilla, which severely injures or kills the plant's growing tips. Feeding damage changes the plant's structure or architecture by preventing new hydrilla stems from reaching the surface of the water column. Despite localized and occasionally severe impacts on hydrilla, none of these insects can cause damage significant enough to provide adequate control

when used alone. Research to identify biocontrol agents for hydrilla continues due to the increasing spread of the species throughout the US, its development of resistance to the herbicide fluridone and the relatively high costs associated with other methods employed to control this weed.

Purple loosestrife (Chapter 15.12)

(see <http://el.ercd.usace.army.mil/aqua/APIS/apishelp.htm> for more information)

Enemy	Type	Origin (date)	Success	Comments
<i>Galerucella californiensis</i>	Beetle	Germany (1992)	Substantial	Widely distributed throughout the range of purple loosestrife in the US
<i>Galerucella pusilla</i>	Beetle	Germany (1992)	Substantial	
<i>Hylobius transversovittatus</i>	Weevil	Germany (1992)	Substantial	
<i>Nanophyes marmoratus</i>	Weevil	France, Germany (1994)	Negligible?	

Two nearly identical *Galerucella* leaf beetles are responsible for most biocontrol of purple loosestrife; in fact, these beetles have reduced purple loosestrife infestations by 90% in several states, especially Oregon and Washington. Larvae feed on buds, leaves and stems of the plants and heavily defoliated plants are often killed by the feeding insects. The life cycle of the beetles is completed in about 6 weeks but there is only one generation per year, with pupation occurring in the soil if it is not continuously flooded. This low rate of reproduction is responsible for the lag time between introduction of the beetles and noticeable effects on the plants. Two weevils – the root-attacking *Hylobius* and seed-attacking *Nanophyes* – also contribute to the successful biocontrol of purple loosestrife. Larvae of *Hylobius* feed and develop in the tap roots and pupation occurs in the upper part of the root. Larvae require 1 to 2 years to complete their development and adults can live for several years. Adults of *Nanophyes* feed on young leaves or flowers and lay their eggs in flower buds. Pupation occurs inside the bud and larvae consume the flower buds; buds then fail to open and drop prematurely from the plant. Although the entire life cycle is completed in about 1 month, there is only 1 generation per year. Leaf-eating *Galerucella* beetles, root-attacking *Hylobius* weevils and seed-attacking *Nanophyes* weevils have only recently been introduced as biocontrol agents on purple loosestrife but appear to be very successful in reducing the growth, occurrence and competitiveness of this emergent weed.

Eurasian watermilfoil (Chapter 15.2)

(see <http://www.invasive.org/eastern/biocontrol/6EurasianMilfoil.html> for more information)

Enemy	Type	Origin (date)	Success	Comments
<i>Acentria ephemerella</i>	Moth	Europe (adventive)	Negligible?	All can cause declines to populations of Eurasian watermilfoil in localized areas of lakes. Results are difficult to predict.
<i>Cricotopus myriophylli</i>	Midge	China (adventive)	Negligible?	
<i>Eurychiopsis lecontei</i>	Weevil	US (native)	Substantial?	

Several insects have been found attacking Eurasian watermilfoil during overseas surveys, but none have been introduced to the US thus far. Recent declines in the abundance of Eurasian watermilfoil in some northern lakes have been attributed to the adventive *Acentria* moth and *Cricotopus* midge, as well as the native *Eurychiopsis* weevil. These insects are widely distributed throughout the range of Eurasian watermilfoil in North America and are found in all areas infested by the weed; as a

result, it is difficult to assess their effectiveness as biocontrol agents. Larvae of the *Acentria* moth feed both in and on stems and leaves, which causes the leaves to drop off the plant. Females have reduced wings and are usually flightless and mating occurs in or on the water surface. Two generations are produced annually and pupae form on the stems. Larvae also feed on a variety of native plants in the absence of Eurasian watermilfoil, so the *Acentria* moth is not a typical biocontrol agent. The *Cricotopus* midge is widely distributed and has been shown to reduce the growth and biomass of Eurasian watermilfoil in laboratory experiments. This midge is not the same species of *Cricotopus* that attacks hydrilla, which suggests these insects may be host specific. It is worth noting that midges rarely feed on living plant tissue and most species typically feed on decaying organic matter. The *Eurychiopsis* weevil is generally considered to be the most important biocontrol agent of Eurasian watermilfoil from an operational perspective even though it is a native insect because this weevil prefers Eurasian watermilfoil over its native natural host. The life cycle of the weevil is completed in about 30 days; adults feed on leaves and stems, whereas larvae are stem borers that consume apical meristems. Feeding damage causes the stems to break apart and heavy feeding by the insects prevents the formation of surface mats. High populations of the *Eurychiopsis* weevil have been associated with declines of populations of Eurasian watermilfoil in some northeastern and midwestern states but fish predation may prevent this weevil from reaching its full biocontrol potential. The *Eurychiopsis* weevil is commercially available and can be purchased to augment existing weevil populations. However, research studying the value of augmenting existing populations with purchased insects has been inconclusive.

Waterlettuce – Chapter 15.8

(see <http://el.erdc.usace.army.mil/aqua/APIS/apishelp.htm> for more information)

Enemy	Type	Origin (date)	Success	Comments
<i>Spodoptera pectinicornis</i>	Moth	Thailand (1990)	Not Established	May be affected by predation by other insects
<i>Neohydronomus affinis</i>	Weevil	Brazil (1987)	Negligible?	

Waterlettuce is a tropical species that is believed to be native to North America and was extirpated (died out) during the Ice Ages, but was reintroduced into Florida in the 16th century. It forms large floating mats similar to those of waterhyacinth in the extreme southern US and populations of waterlettuce often increase as waterhyacinth populations decline. Waterlettuce is a public health issue in Florida, where larvae of disease-causing *Mansonia* mosquitoes (Chapter 5) attach to the extensive feathery roots to obtain oxygen. Two insects have been released as biocontrol agents of waterlettuce but only the *Neohydronomus* weevil has become established.

Adults and larvae of the *Neohydronomus* weevil feed on the leaves, crown and newly emerging shoots of waterlettuce and the characteristic "shot hole" appearance of leaves indicates high weevil densities. Feeding by multiple larvae destroys the spongy leaf bases, which causes plants to lose buoyancy. The life cycle of the *Neohydronomus* weevil is completed in 3 to 4 weeks. The weevil has not contributed to long-term suppression of the plant in the US, but has provided successful biocontrol of waterlettuce in other countries. It is thought that the *Neohydronomus* weevil is heavily preyed upon by imported fire ants in Florida; if true, this provides an interesting example of an exotic invader controlling a valuable potential biocontrol agent.

Giant salvinia (Chapter 15.9)

(see <http://el.ercd.usace.army.mil/aqua/APIS/apishelp.htm> for more information)

Enemy	Type	Origin (date)	Success	Comments
<i>Cyrtobagous salviniae</i>	Weevil	Brazil? (adventive)	Negligible, Substantial	Provides good control of common salvinia in FL but not elsewhere.
<i>Cyrtobagous salviniae</i>	Weevil	Brazil (2001)	Substantial?	Effects of 2001 introduction on giant salvinia are still being evaluated

The *Cyrtobagous* weevil is the only insect that has been released as a biocontrol agent of giant salvinia. Adventive weevils that were discovered in Florida in 1960 are used to control common salvinia (*Salvinia minima*), whereas weevils released in 2001 from a Brazilian population are used as biocontrol agents for giant salvinia. The entire life cycle of the *Cyrtobagous* weevil takes about 46 days. Adults feed on leaf buds and leaves and larvae tunnel inside the plant, killing leaves and rhizomes. Attacked plants turn brown and eventually lose buoyancy. *Cyrtobagous* weevils from Australia are currently of great interest to researchers and have been introduced as biocontrol agents for giant salvinia, but it is too early to determine the effectiveness of these weevils in the US.

Melaleuca (*Melaleuca quinquenerva*)

Enemy	Type	Origin (date)	Success	Comments
<i>Oxyops vitiosa</i>	Weevil	Australia (1997)	Substantial	Not established in permanently flooded sites due to inability to complete life cycle. Also see http://edis.ifas.ufl.edu/document_in172
<i>Boreioglycaspis melaleucae</i>	Psyllid	Australia (2002)	Substantial	
<i>Fergusonina turneri</i>	Fly	Australia (2005)	Not Established	
<i>Lophodiplosis trifida</i>	Fly	Australia (2008)	Negligible	Establishment confirmed http://tame.ifas.ufl.edu/photo_gallery/bioco ntrol/stem-gall-fly.shtml

Melaleuca is a locally invasive plant that occurs only in south Florida and the Everglades and was introduced multiple times during the early 1900s. The species was used as an ornamental tree and was planted in marshes to drain wetlands. Melaleuca typically grows in dense, impenetrable stands and can attain a height over 50 feet. Four insects have been released as biocontrol agents of melaleuca but only three have become established.

The *Oxyops* weevil and the *Boreioglycaspis* psyllid were released in 1997 and 2002, respectively, and are widely established on melaleuca in south Florida. Damage to the tree is caused primarily by the immature stages of these insects. The slug-like weevil larvae feed on newly expanding leaves; psyllid nymphs attack older leaves and woody stems in addition to new leaves and the psyllid can kill newly emerged seedlings as well. These two insects complement each other well; the psyllid is able to complete its development entirely in the tree canopy under flooded conditions that prevent establishment of the weevil, which must pupate in the soil. Extensive leaf damage from both insects causes melaleuca to divert resources to the production of new foliage instead of flowers. The life cycle of the weevil is completed in about 3 months, whereas a new psyllid generation is produced in 6 weeks. The *Oxyops* weevil and the *Boreioglycaspis* psyllid have contributed to the substantial

biocontrol of melaleuca. The *Lophodiplosis* gall-forming fly was released in 2008 and has apparently become established; however, it is too early to assess its impact on melaleuca.

Summary

The use of insects as biological control agents for aquatic weeds has yielded mixed results, which is typical and expected of biocontrol programs. A number of aquatic weeds – including alligatorweed, purple loosestrife and melaleuca – are being successfully controlled by insects released as biocontrol agents for these species. Control of other aquatic weeds – including waterhyacinth, hydrilla, Eurasian watermilfoil, waterlettuce and giant salvinia – has been less successful. Multiple factors play a role in the failure of some biocontrol agents to reach their full potential. For example, the *Neohydronomus* weevil has provided successful biocontrol of waterlettuce in other countries, but has failed to control waterlettuce in Florida, possibly due to predation of the weevil by imported fire ants. Biocontrol can be an effective tool in the aquatic weed manager's arsenal since host-specific biocontrol agents allow management of populations of weedy species while leaving nontarget native plants unharmed. Therefore, it is important that researchers continue to identify and evaluate biocontrol agents so that the successes realized in the control of alligatorweed, purple loosestrife and melaleuca can be duplicated in other weedy aquatic species. A major factor that limits the utility of biocontrol is that unless a potential biocontrol agent is species-specific, it cannot be introduced into the US. Therefore, it is unlikely that biocontrol alone can control all the invasive aquatic weeds in the US.

For more information:

- Harley KLS and IW Forno. 1992. Biological control of weeds: a handbook for practitioners and students. Inkata Press, Melbourne, Australia.

Photo and illustration credits:

Page 59: Alligatorweed flea beetle; Gary Buckingham, USDA-ARS
Page 60: Graph; from Harley and Forno, 1992

